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• 综述 •

烟曲霉耐药机制及中药单体干预作用的研究进展*

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【摘要】 侵袭性曲霉病(invasive aspergillosis, IA)是感染曲霉菌引发的疾病,烟曲霉是常见的引起IA和过敏性疾病的曲霉属物种,主要通过掩蔽其表面分子、调节免疫反应等级制来逃避和适应宿主,临床大剂量使用抗真菌药物,导致耐药菌株数量在不断增加,给临床治疗带来困难。近年来,中药因其毒副作用小、耐药率低等原因,在抗真菌治疗方面有一定优势。因此研究烟曲霉的耐药机制并探讨中药单体干预作用,对寻找烟曲霉耐药机制相关靶点及中药抗真菌研究、减少耐药菌株形成具有重要意义。

【关键词】 烟曲霉;耐药机制;中药单体;综述

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Research progress on drug resistance mechanism of *Aspergillus fumigatus* and the intervention effect of herbal monomer

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【Abstract】 Invasive aspergillosis (IA) refers to a variety of diseases caused by *Aspergillus*, and *Aspergillus fumigatus* is the most common species of *Aspergillus* causing IA and allergic diseases. At the same time, the clinical use of large doses of antifungal drugs, resulting in the number of drug-resistant strains is increasing, bringing difficulties to clinical treatment. In recent years, traditional Chinese medicine has had some advantages in antifungal treatment because of its low toxic side effects and low drug resistance rate. Therefore, studying the drug resistance mechanism of *A. fumigatus* and exploring the effect of the monomeric intervention of traditional Chinese medicine is important for finding the targets related to the drug resistance mechanism of *A. fumigatus* and the antifungal research of traditional Chinese medicine and reducing the formation of drug-resistant strains.

【Key words】 *Aspergillus fumigatus*; drug resistance mechanism; monomer of traditional Chinese medicine; review

***曲霉属是一类分布广泛的真菌属,包括300多种不同的物种,曲霉感染称为侵袭性曲霉病(invasive aspergillosis, IA),包括导致过敏反应的感染,如过敏性曲霉鼻窦炎^[1]或过敏性支气管肺曲霉病^[2],皮肤感染导致皮肤曲霉病^[3],曲菌瘤和慢性疾病如慢性肺曲菌病^[4]。IA主要影响免疫功能低下患者,例如癌症患者、接受皮质醇治疗和器官移植者^[3,5],但少数情况下亦可有免疫正常患者^[6-7]。IA已成为现代真菌学的主要临床问题之一,其中烟曲霉是最常见的引起IA和过敏性疾病的曲霉属物种。近些年来,烟曲霉耐药菌株数量不断增加,抗真菌药物的疗效也随之降低,中药单体可以通过多途径抑制烟曲霉生物膜形成、减少烟曲霉毒力,改善病理损伤。现就烟曲霉耐药机制及中药单体干预相关研究予以综述,以期为烟曲霉耐药的中医药研究提供新途径。

1 烟曲霉耐药机制

近年来,随着免疫功能受损患者的增多,IA的发病率随之增加,尽管已经对该病早期诊断和治疗方面研究有所进展,但IA的死亡率仍然较高。未经治疗时的死亡率为80%~95%^[8],用伏立康唑(VRC)或两性霉素B(Amphotericin B, AmB)治疗12周后的死亡率分别为29.2%和42.1%^[9]。其原因包括宿主免疫缺陷难以抵御真菌、IA早期诊断困难以及耐

药菌株的出现。烟曲霉作为最常引起IA的曲霉属菌,研究其耐药机制有助于临床治疗IA,降低患者死亡率。目前常用抗真菌药物有唑类、多烯类和棘白菌素类。

1.1 増类 増类通过抑制甾醇14 α -去甲基酶(steryl 14 α -demethylase, Cyp51)蛋白引起羊毛甾醇积聚及麦角甾醇缺乏从而发挥固有的抗真菌活性,在此基础上,烟曲霉主要耐药机制可分为,Cyp51蛋白突变、外排泵上调、细胞应激反应、HMG-CoA编码基因突变和生物膜形成。除此之外,细胞色素氧化酶也被认为是烟曲霉的耐药机制之一^[10-11]。

1.1.1 Cyp51蛋白 増类耐药机制的研究中,最常检测到的突变与靶蛋白Cyp51A有关。麦角甾醇是微生物细胞膜的重要组成成分,对确保细胞活力、膜的流动性、膜结合酶的活性、

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膜的完整性以及细胞物质运输等起着重要作用^[12],该酶参与麦角甾醇生物合成和甾醇代谢,在烟曲霉中起重要作用。Cyp51A 拥有同源蛋白 Cyp51B,这两种蛋白对于烟曲霉生长不是必须的^[13-14],但失活是致命的^[15],唑类与 Cyp51 蛋白相互作用并抑制 Cyp51 蛋白^[16],反过来又降低麦角甾醇含量并破坏细胞中的甾醇代谢^[17],从而发挥杀菌作用。

1.1.1.1 Cyp51 蛋白单点突变 Cyp51A 基因由于蛋白内氨基酸置换所引起的单点突变是烟曲霉主要耐药机制,该突变会改变 Cyp51 的结构、稳定性和功能性,进而阻碍底物识别,最终导致唑耐药^[16,18]。迄今为止,由 G54、Y121、G138、P216、F219、M220、A284、Y431、G432、G434 和 G448 位氨基酸取代组成的烟曲霉 Cyp51A 已被发现与产生唑耐药有关^[19-33]。在没有 Cyp51B 的情况下,Cyp51A 蛋白表达增加,反之亦然,表明一种蛋白质在蛋白质水平上补偿另一种蛋白质缺失的能力。在唑类耐药性背景下,Cyp51A 对唑类敏感性的影响大于 Cyp51B 的影响^[34]。Handelman 等^[35]研究发现在耐药临床分离株中鉴定出的 G457S Cyp51B 突变的引入会导致野生型受体菌株中对 VRC 产生唑耐药。

1.1.1.2 Cyp51 过表达 另一种被认为导致烟曲霉产生耐药性的机制是 Cyp51 的过表达。当转录因子和转录激活物接触到唑类化合物进而导致 Cyp51 转录上调。在烟曲霉 Cyp51A 启动子区存在串联重复序列 (tandem repeats, TRs): 34 bp (TR34)、46 bp (TR46) 或 53 bp (TR53) 的情况下,该基因的表达增强^[36-39]。

由于甾醇生物合成受到高度调控,许多编码该途径酶的基因在其启动子区含有甾醇调节结合元件^[40]。与烟曲霉 Cyp51 启动子结合并调节其表达的转录因子 (transcription factors, TF),如 SrbA、HapE 和 AtrR。CCAAT 结合复合物 (CCAAT-binding complex, CBC) 是重要的一般转录调节因子,CBC-HapX 复合物是铁稳态的主要调节剂,Alexander Kühbacher 等发现破坏铁调节剂 HapX 的结合会增加 cyp51A 表达^[41]。此外,细胞色素 b₅-CybE 可调节烟曲霉 Cyp51A 的转录水平,CybE 的缺失导致 Cyp51A 的代偿性上调^[42]。

1.1.2 外排转运蛋白 多药物外排泵由跨膜蛋白组成,其介导抗菌分子或有毒化合物和内源性代谢物向细胞外空间的主动排出^[43]。因此,外排活性是耐药性和真菌存活的决定性因素。目前有两种已知的外排转运蛋白超家族:ATP 结合盒 (ATP-binding cassette, ABC) 和主要协同蛋白超家族 (major facilitator superfamily, MFS) 转运蛋白。烟曲霉基因组中鉴定出 45 个 ABC 和 275 个 MFS 转运蛋白^[44],但是只有少数被鉴定为药物蛋白。ATP 结合盒是由两个跨膜结构域和两个细胞质核苷酸结合结构域组成,通过 ATP 水解产生的能量将底物排出,MFS 超家族利用质膜上质子梯度动力排出药物^[45-46]。

烟曲霉含有至少 49 个编码 ABC 转运蛋白的基因^[47]。其中,Cdr1B 被鉴定为在唑类耐药菌株中过表达,并且删除此类菌株中的 Cdr1B 基因导致对伊曲康唑的敏感性增加^[48],其他删除 Cdr1B 基因的菌株显示出唑类超敏反应^[49]。关于 MFS 转运蛋白目前研究仅有 mfsA、mfsB 和 mfsC 被证实在烟曲霉 唑类药物暴露期间上调^[50]。

1.1.3 细胞应激反应 应激相关蛋白在应激条件下通过这些

蛋白质控制基因表达从而产生耐唑性。热休克蛋白 90 (heat shock protein 90, Hsp90) 是一种重要的分子伴侣蛋白,通过与其催化亚基相互作用来激活钙调磷酸酶,以调节应激反应,包括烟曲霉中唑诱导的应激。抑制 Hsp90 或钙调神经磷酸酶会增加烟曲霉对唑类的敏感性^[51-52]。

损伤抗性蛋白家族 (the damage resistance protein, Dap) 同样会对唑类产生应激反应,该家族由 DapA、DapB、DapC 组成,有研究表明,DapA 的缺失增加了唑的敏感性,因此其可能与唑抗性有关^[53-54]。

Ca²⁺ 作为第二信使 在细胞功能调节中起重要作用,由 Ca²⁺ 通道蛋白、钙泵、Ca²⁺ 转运蛋白和许多相关蛋白质组成的钙信号通路在真菌中起着调节作用。Crz1 是钙信号通路下游的转录因子,参与调节细胞存活、离子稳态、感染结构发育、细胞壁完整性和毒力。Crz1 的缺失会增加对唑类用药物的敏感性^[55]。

1.1.4 HMG-CoA 编码基因突变 研究发现 3-羟基-3-甲基戊二酰辅酶 A (HMG-CoA) 还原酶编码基因 Hmg1 的突变导致对所有临床可用的唑类药物的耐药性显著增加。Gonzalez 等^[56]实验证明唑耐药与 Cyp51B (G457S) 和 Hmg1 (F390L) 的修饰有关。在烟曲霉中,除了 Hmg1 外,还存在第二种 HMG-CoA 还原酶 Hmg2,目前尚未有研究证明其与耐药有关,可能是未来研究的新方向^[57]。

1.1.5 生物膜形成 生物膜指的是附在物体表面被细菌胞外大分子包裹的有组织的细菌群体。生物膜形成是烟曲霉耐药的重要机制,生物膜使得烟曲霉对抗真菌药物的敏感性降低。成熟生物膜中达到的细胞密度可能会阻碍药物渗透,因为它的疏水性与菌丝紧密结合^[58]。

1.1.6 细胞色素氧化酶 研究发现一种新的细胞色素 C 氧化酶 cox7c,其缺失或突变明确导致对唑类、多烯类和烯丙胺药物的耐药性^[59]。

1.2 多烯类 多烯是第一种上市广谱抗真菌药物,但是由于其显著的肝肾毒性使得 AmB 等多烯类药物颇具争议。多烯抗真菌的关键是与麦角甾醇的结合^[60]。该类药物与真菌细胞膜上的麦角甾醇结合,使细胞膜上形成微孔,改变细胞膜的通透性,从而引起细胞内小分子和离子外渗;随着其浓度的增高,大分子也可通过细胞膜外渗,导致细胞内成分不可逆的丢失,而致真菌死亡^[61]。AmB 暴露会诱导烟曲霉中细胞内活性氧 (reactive oxygen species, ROS) 的产生和积累,从而导致氧化损伤^[62]。

多烯耐药性是由麦角甾醇生物合成基因的功能缺失突变引起的(特别是在曲霉属和念珠菌属),在曲霉属中,只有土曲霉对 AmB 天然耐药^[63],其余发生耐药反应比较少见,而烟曲霉有 43%~96.4% 的概率产生 AmB 抗性^[64-65]。烟曲霉多烯类耐药机制目前尚不明确,Fan 等^[66]研究确定了 34 个与 AmB MIC 差异相关的候选单核苷酸多态性 (single-nucleotide polymorphisms, SNP),包括 18 个基因间变异,14 个错义变异,1 个同义变异和 1 个非编码转录本变异,有助于快速筛选烟曲霉抗 AMB 基因。

1.3 棘白菌素类 真菌细胞壁成分包括 1,3-β-D-葡聚糖、1,4-β-D-葡聚糖、1,6-β-D-葡聚糖、α-葡聚糖、甲壳素、甘露聚糖和多种糖蛋白,葡聚糖是细胞壁的重要结构成分,在保护环境、控制

渗透压、菌丝形态发生和宿主组织中的侵袭性方面起着至关重要的作用^[67]。并且,1,3-β-D-葡聚糖在动物细胞中不存在,因此它是抗真菌抗生素的优选靶标^[68]。1,3-β-D-葡聚糖合成酶是由至少两个亚基组成的跨膜异构糖基转移酶,其中Fks1p亚基(由Fks1,Fks2和Fks3基因编码)具有催化功能,而Rho1p亚基(属于GTP酶家族)具有调节功能。棘白菌素与该酶的Fks1p亚基非竞争性结合抑制其活性^[3,69],从而抑制1,3-β-D-葡聚糖合成酶,影响烟曲霉细胞壁合成,使其生长过程中细胞壁葡聚糖缺乏、渗透压失常而最终导致细胞溶解^[70]。在3大抗真菌药物类别中,棘白菌素对曲霉菌属药效最低,但是对比唑类和AmB,棘白菌素的优势在于较少的药物相互作用和相关毒性。

1.3.1 葡聚糖合酶催化亚基突变 棘白菌素耐药性的研究涉及葡聚糖合酶(FKS)亚基的突变。编码该亚基的三个基因是已知的:FKS1、FKS2和FKS3。烟曲霉的突变发生在葡聚糖合酶的*A. fumigatus* FKS1基因中,随着几丁质产生的增加,烟曲霉对棘白菌素产生耐药性^[71]。在Fks1中具有S678Y或S678P突变的实验室烟曲霉菌株具有与三种棘白菌素类药物相关表型的耐药性,从而证明其耐药^[72-73]。

1.3.2 钙调磷酸酶途径 曲霉菌属在适应棘白菌素后会产生应激补偿机制,这种机制被称为矛盾效应(Paradoxical effect,PE)^[74],指的是真菌随着药物浓度升高敏感性下降,甚至产生耐药。热休克蛋白90(Hsp90)和70(Hsp70)是重要的分子伴侣,当卡泊芬净达到一定浓度作用于烟曲霉产生应激反应时,Hsp90和Hsp70可能通过协同作用以控制钙调磷酸酶途径产生PE。PE的关键适应机制与1,3-β-葡聚糖合酶活性的恢复有关。1,3-β-葡聚糖的合成通过1,3-β-葡聚糖合酶复合物在细胞膜上进行,0.5 μg/mL浓度下,Fks1从菌丝尖端错误定位到液泡。然而,仅连续暴露于4 μg/mL卡泊芬净48 h就会使菌丝形态恢复正常,Fks1重新定位到菌丝尖端。但是用两种浓度卡泊芬净处理后,Rho1仍保留在菌丝尖端,用法尼醇处理烟曲霉会导致生长菌丝顶端的Rho1和Rho3定位错误,通过这种途径阻断细胞壁完整性^[75],说明Rho1对于PE是必需的^[76]。

1.3.3 细胞壁完整性 烟曲霉生物膜内的菌丝含有细胞外基质(extracellular matrix, ECM)和多糖半乳糖氨基半乳聚糖(galactosaminogalactan,GAG),GAG是烟曲霉生物膜的主要粘附因子以及免疫调节化合物,当烟曲霉处于缺氧环境时,丙氨酸氨基转移酶(alanine aminotransferase,alaA)的mRNA水平大幅增加,alaA的缺失导致GAG粘附功能的改变,进而导致细胞壁变化和生物膜对棘白菌素的敏感性增加^[77]。卡泊芬净导致的细胞壁应激反应会诱导生物膜形成,且调节GAG和细胞壁多糖的生物合成的转录因子SomA下调会导致严重的生物膜形成缺陷和对细胞壁应激源的超敏反应^[78]。

2 中药单体对烟曲霉的干预作用

目前中药单体抗真菌的研究备受关注,中药单体具有获取便利,耐药率低,毒副作用小等优点。对于治疗烟曲霉耐药具有独特优势,中药单体可以靶向烟曲霉生物膜,破坏细胞壁的连续性,减弱烟曲霉毒力,改善病理损伤等,还可与临床常用抗真菌药物联用,防止耐药。

2.1 肉桂醛 肉桂醛又叫桂醛,主要是从桂皮、桂叶和桂枝中提取出来的一种芳香醛类有机化合物,对细菌和真菌具有较强

的抑制作用,并具有抗生物膜活性^[79-80],对临床分离的烟曲霉具有抗菌活性^[81],邬丽红等^[82]研究表明肉桂醛作用于已经成熟的烟曲霉菌生物膜后,生物膜的致密结构变松散,菌丝表面及菌丝之间胞外基质被清除,且在高浓度时部分菌丝被裂解,提示肉桂醛防止烟曲霉生物膜的形成或者破坏已经形成的生物膜结构,使得药物能够克服生物膜屏障渗透到烟曲霉细胞内达到有效浓度而发挥抗菌作用。然而肉桂醛在水溶液中的溶解度差、不稳定和挥发性^[83]。因此,应在未来的研究中进一步考虑使用不同的给药系统以解决这些局限性。

2.2 柠檬醛 柠檬醛是中药山苍子的主要成分,属萜类化合物,龙凯等^[84]研究发现柠檬醛能够在体外有效抑制烟曲霉。罗闳丹等^[85]研究发现中、高剂量柠檬醛能够有效阻止烟曲霉孢子在肺组织的萌发和菌丝生长,且与烟曲霉毒力密切相关的烟灰色色素明显减弱,进而改善IPA模型小鼠各器官的病理损伤。

2.3 大蒜素 大蒜素(二烯丙基硫代硫酸盐)是新鲜压碎的大蒜的活性化合物之一。大蒜素具有抗菌、抗炎、抗血栓、抗动脉粥样硬化、降血脂和抗癌等多种生物活性。大蒜提取物对新生隐球菌^[86]、念珠菌和曲霉菌均具有体外抗菌活性^[87]。Shadkhan等^[88]研究发现纯大蒜素可能是一种有效的体外杀菌剂,时间杀伤研究表明,纯大蒜素在给药后2~12 h内发挥杀菌活性,大蒜素治疗显著延长了感染小鼠的存活时间,以前的研究认为即使是最有效的大蒜素治疗也不如AmB治疗成功,但是通过化学合成的大蒜素在直接接触时,通过气相对选定对真菌具有体外杀菌活性,其浓度与临床使用的AmB相当^[89]。大蒜素缺点主要是在血液、溶剂和模拟生理液中的半衰期很短(50 min)^[90],且在培养过程中,大蒜素对哺乳动物的毒性明显高于曲霉属,因此需要进行更多的研究以找到最佳的治疗方式。

2.4 小檗碱 小檗碱是从几种草药中分离出来的生物碱,具有多种药理作用,包括抗菌、抗糖尿病和抗癌活性。高磊^[91]研究发现小檗碱使菌丝和孢子形态发生畸变:透射电子显微镜观察可见小檗碱能破坏烟曲霉细胞壁的连续性,使细胞膜部分缺失,细胞器内容物减少,AmB与小檗碱联合用药可以通过降低AmB药量的方式降低AmB毒副作用。

3 小结

本文探讨了烟曲霉的3大类抗菌药物耐药机制,以及中药单体干预作用。目前,由于其耐药机制复杂,还需对抗真菌药物调节因子、多种抗真菌药物相互作用、生物膜形成机制、耐药靶点、转录因子等进行探索。生物膜形成是烟曲霉主要耐药原因之一。肉桂醛、柠檬醛能够调节生物膜形成,破坏细胞壁的连续性,增加烟曲霉对抗真菌药物敏感性,同时降低烟曲霉毒力,改善宿主病理损伤与抗真菌药物联用可以降低毒副作用。大蒜素可制成雾化剂与AmB联用治疗肺真菌病。中药单体主要的功能不是杀死烟曲霉而是抑制其产生耐药性和辅助抗真菌药物,可为研发治疗由耐药菌株引起的感染,以抑制耐药的产生的新药理论基础。

【参考文献】

- [1] Chakrabarti A, Kaur H. Allergic *Aspergillus rhinosinusitis* [J]. J Fungi (Basel), 2016, 2(4):32.

- [2] Patel G, Greenberger PA. Allergic bronchopulmonary aspergillosis[J]. *Allergy Asthma Proc*, 2019, 40(6):421-424.
- [3] Patterson TF, Thompson GR 3rd, Denning DW, et al. Practice guidelines for the diagnosis and management of aspergillosis; 2016 update by the infectious diseases society of America[J]. *Clin Infect Dis*, 2016, 63(4):e1-e60.
- [4] Kosmidis C, Denning DW. The clinical spectrum of pulmonary aspergillosis[J]. *Thorax*, 2015, 70(3):270-277.
- [5] Desoubeaux G, Bailly E, Chandenier J. Diagnosis of invasive pulmonary aspergillosis; updates and recommendations[J]. *Med Mal Infect*, 2014, 44(3):89-101.
- [6] Mohammed AP, Dhunpath P, Chiluka R, et al. An unusual case of invasive aspergillosis in an immunocompetent individual[J]. *BMJ Case Rep*, 2015(29):1757-1790.
- [7] Cheon S, Yang MK, Kim CJ, et al. Disseminated aspergillosis in the immunocompetent host: A case report and literature review [J]. *Mycopathologia*, 2015, 180(3-4):217-222.
- [8] Kontoyiannis DP, Bodey GP. Invasive aspergillosis in 2002: an update[J]. *Eur J Clin Microbiol Infect Dis*, 2002, 21(3):161-172.
- [9] Ajmal S, Mahmood M, Abu Saleh O, et al. Invasive fungal infections associated with prior respiratory viral infections in immunocompromised hosts [J]. *Infection*, 2018, 46(4):555-558.
- [10] Hagiwara D, Watanabe A, Kamei K, et al. Epidemiological and genomic landscape of azole resistance mechanisms in *Aspergillus fungi*[J]. *Front Microbiol*, 2016(7):1382.
- [11] Wei X, Zhang Y, Lu L. The molecular mechanism of azole resistance in *Aspergillus fumigatus*: from bedside to bench and back[J]. *J Microbiol (Seoul, Korea)*, 2015, 53(2):91-99.
- [12] 韩庆雪,闻建平,冯国龙.麦角固醇工业化提取工艺研究[J].河北化工,2007,8(02):42-43.
- [13] Garcia-Effron G, Mellado E, Gomez-Lopez A, et al. Differences in interactions between azole drugs related to modifications in the 14-alpha sterol demethylase gene (cyp51A) of *Aspergillus fumigatus*[J]. *Antimicrob Agents Chemother*, 2005, 49 (5): 2119-2121.
- [14] Mellado E, Garcia-Effron G, Buitrago MJ, et al. Targeted gene disruption of the 14-alpha sterol demethylase (cyp51A) in *Aspergillus fumigatus* and its role in azole drug susceptibility[J]. *Antimicrob Agents Chemother*, 2005, 49(6):2536-2538.
- [15] Hu W, Sillaots S, Lemieux S, et al. Essential gene identification and drug target prioritization in *Aspergillus fumigatus* [J]. *PLoS Pathog*, 2007, 3(3):e24.
- [16] Warrilow AG, Parker JE, Price CL, et al. *In vitro* biochemical study of CYP51-mediated azole resistance in *Aspergillus fumigatus*[J]. *Antimicrob Agents Chemother*, 2015, 59 (12): 7771-7778.
- [17] Alcazar-Fuoli L, Mellado E, Garcia-Effron G, et al. Ergosterol biosynthesis pathway in *Aspergillus fumigatus*[J]. *Steroids*, 2008, 73(3):339-347.
- [18] Liu M, Zheng N, Li D, et al. cyp51A-based mechanism of azole resistance in *Aspergillus fumigatus*: Illustration by a new 3D Structural Model of *Aspergillus fumigatus* CYP51A protein [J]. *Med Mycol*, 2016, 54(4):400-408.
- [19] Mann PA, Parmegiani RM, Wei SQ, et al. Mutations in *Aspergillus fumigatus* resulting in reduced susceptibility to posaconazole appear to be restricted to a single amino acid in the cytochrome P450 14alpha-demethylase[J]. *Antimicrob Agents Chemother*, 2003, 47(2):577-581.
- [20] Nascimento AM, Goldman GH, Park S, et al. Multiple resistance mechanisms among *Aspergillus fumigatus* mutants with high-level resistance to itraconazole[J]. *Antimicrob Agents Chemother*, 2003, 47(5):1719-1726.
- [21] Krishnan NS, Wu W, Cutright JL, et al. *In vitro-in vivo* correlation of voriconazole resistance due to G448S mutation (cyp51A gene) in *Aspergillus fumigatus*[J]. *Diagn Microbiol Infect Dis*, 2012, 74(3):272-277.
- [22] Bader O, Weig M, Reichard U, et al. cyp51A-Based mechanisms of *Aspergillus fumigatus* azole drug resistance present in clinical samples from Germany[J]. *Antimicrob Agents Chemother*, 2013, 57(8):3513-3517.
- [23] Camps SM, van der Linden JW, Li Y, et al. Rapid induction of multiple resistance mechanisms in *Aspergillus fumigatus* during azole therapy: a case study and review of the literature[J]. *Antimicrob Agents Chemother*, 2012, 56(1):10-16.
- [24] Kidd SE, Goeman E, Meis JF, et al. Multi-triazole-resistant *Aspergillus fumigatus* infections in Australia[J]. *Mycoses*, 2015, 58(6):350-355.
- [25] Alanio A, Sitterl E, Liance M, et al. Low prevalence of resistance to azoles in *Aspergillus fumigatus* in a French cohort of patients treated for haematological malignancies[J]. *J Antimicrob Chemother*, 2011, 66(2):371-374.
- [26] Lescar J, Meyer I, Akshita K, et al. *Aspergillus fumigatus* harbouring the sole Y121F mutation shows decreased susceptibility to voriconazole but maintained susceptibility to itraconazole and posaconazole[J]. *J Antimicrob Chemother*, 2014, 69(12):3244-3247.
- [27] Wiederhold NP, Gil VG, Gutierrez F, et al. First detection of TR34 L98H and TR46 Y121F T289A Cyp51 mutations in *Aspergillus fumigatus* isolates in the United States[J]. *J Clin Microbiol*, 2016, 54(1):168-171.
- [28] Mellado E, Garcia-Effron G, Alcazar-Fuoli L, et al. Substitutions at methionine 220 in the 14alpha-sterol demethylase (Cyp51A) of *Aspergillus fumigatus* are responsible for resistance *in vitro* to azole antifungal drugs[J]. *Antimicrob Agents Chemother*, 2004, 48(7):2747-2750.
- [29] Howard SJ, Cerar D, Anderson MJ, et al. Frequency and evolution of Azole resistance in *Aspergillus fumigatus* associated with treatment failure[J]. *Emerg Infect Dis*, 2009, 15(7):1068-1076.
- [30] Bellete B, Raberin H, Morel J, et al. Acquired resistance to voriconazole and itraconazole in a patient with pulmonary aspergillosis[J]. *Med Mycol*, 2010, 48(1):197-200.
- [31] Bueid A, Howard SJ, Moore CB, et al. Azole antifungal resistance in *Aspergillus fumigatus*; 2008 and 2009[J]. *J Antimicrob Chemother*, 2010, 65(10):2116-2118.
- [32] Snelders E, Karawajczyk A, Schaftenaar G, et al. Azole resistance profile of amino acid changes in *Aspergillus fumigatus* CYP51A based on protein homology modeling[J]. *Antimicrob Agents Chemother*, 2010, 54(6):2425-2430.
- [33] Albarraq AM, Anderson MJ, Howard SJ, et al. Interrogation of

- related clinical pan-azole-resistant *Aspergillus fumigatus* strains; G138C, Y431C, and G434C single nucleotide polymorphisms in cyp51A, upregulation of cyp51A, and integration and activation of transposon Atf1 in the cyp51A promoter[J]. *Antimicrob Agents Chemother*, 2011, 55(11):5113-5121.
- [34] Roundtree MT, Juvvadi PR, Shwab EK, et al. *Aspergillus fumigatus* Cyp51A and Cyp51B proteins are compensatory in function and localize differentially in response to antifungals and cell wall inhibitors[J]. *Antimicrob Agents Chemother*, 2020, 64(10):e00735-20.
- [35] Handelman M, Meir Z, Scott J, et al. Point mutation or overexpression of *Aspergillus fumigatus* cyp51B, encoding lanosterol 14 α -sterol demethylase, leads to triazole resistance[J]. *Antimicrob Agents Chemother*, 2021, 65(10):e0125221.
- [36] Mellado E, Garcia-Effron G, Alcázar-Fuoli L, et al. A new *Aspergillus fumigatus* resistance mechanism conferring in vitro cross-resistance to azole antifungals involves a combination of cyp51A alterations[J]. *Antimicrob Agents Chemother*, 2007, 51(6):1897-1904.
- [37] Snelders E, Karawajczyk A, Verhoeven RJ, et al. The structure-function relationship of the *Aspergillus fumigatus* cyp51A L98H conversion by site-directed mutagenesis: the mechanism of L98H azole resistance[J]. *Fungal Genet Biol*, 2011, 48(11):1062-1070.
- [38] Snelders E, Camps SM, Karawajczyk A, et al. Genotype-phenotype complexity of the TR46/Y121F/T289A cyp51A azole resistance mechanism in *Aspergillus fumigatus*[J]. *Fungal Genet Biol*, 2015, 82:129-135.
- [39] Hodiamont CJ, Dolman KM, Ten Berge IJ, et al. Multiple-azole-resistant *Aspergillus fumigatus* osteomyelitis in a patient with chronic granulomatous disease successfully treated with long-term oral posaconazole and surgery[J]. *Med Mycol*, 2009, 47(2):217-220.
- [40] Dhingra S, Cramer RA. Regulation of Sterol Biosynthesis in the human fungal pathogen *Aspergillus fumigatus*: opportunities for therapeutic development[J]. *Front Microbiol*, 2017(8):92.
- [41] Khbacher A, Peiffer M, Hortschansky P, et al. Azole resistance-associated regulatory motifs within the promoter of cyp51A in *Aspergillus fumigatus*[J]. *Microbiol Spectr*, 2022, 10(3):e0120922.
- [42] Misslinger M, Gsaller F, Hortschansky P, et al. The cytochrome b(5) CybE is regulated by iron availability and is crucial for azole resistance in *A. fumigatus*[J]. *Metallomics*, 2017, 9(11):1655-1665.
- [43] Rajendran R, Mowat E, McCulloch E, et al. Azole resistance of *Aspergillus fumigatus* biofilms is partly associated with efflux pump activity[J]. *Antimicrob Agents Chemother*, 2011, 55(5):2092-2097.
- [44] Meneau I, Coste AT, Sanglard D. Identification of *Aspergillus fumigatus* multidrug transporter genes and their potential involvement in antifungal resistance[J]. *Med Mycol*, 2016, 54(6):616-627.
- [45] Chamilos G, Kontoyiannis DP. Update on antifungal drug resistance mechanisms of *Aspergillus fumigatus*[J]. *Drug Resist Updat*, 2005, 8(6):344-358.
- [46] Law CJ, Maloney PC, Wang DN. Ins and outs of major facilitator superfamily antiporters[J]. *Annu Rev Microbiol*, 2008(62):289-305.
- [47] Leli vre L, Groh M, Angebault C, et al. Azole resistant *Aspergillus fumigatus*: an emerging problem[J]. *Med Mal Infect*, 2013, 43(4):139-145.
- [48] Fraczek MG, Bromley M, Buied A, et al. The cdr1B efflux transporter is associated with non-cyp51a-mediated itraconazole resistance in *Aspergillus fumigatus*[J]. *J Antimicrob Chemother*, 2013, 68(7):1486-1496.
- [49] Paul S, Diekema D, Moye-Rowley WS. Contributions of *Aspergillus fumigatus* ATP-binding cassette transporter proteins to drug resistance and virulence[J]. *Eukaryot Cell*, 2013, 12(12):1619-1628.
- [50] da Silva Ferreira ME, Malavazi I, Savoldi M, et al. Transcriptome analysis of *Aspergillus fumigatus* exposed to voriconazole [J]. *Curr Genet*, 2006, 50(1):32-44.
- [51] Cowen LE. Hsp90 orchestrates stress response signaling governing fungal drug resistance[J]. *PLoS Pathog*, 2009, 5(8):e1000471.
- [52] Lamoth F, Juvvadi PR, Soderblom EJ, et al. Identification of a key lysine residue in heat shock protein 90 required for azole and echinocandin resistance in *Aspergillus fumigatus*[J]. *Antimicrob Agents Chemother*, 2014, 58(4):1889-1896.
- [53] Song J, Zhai P, Lu L. Damage resistance protein (Dap) contributes to azole resistance in a sterol-regulatory-element-binding protein SrbA-dependent way[J]. *Appl Microbiol Biotechnol*, 2017, 101(9):3729-3741.
- [54] Song J, Zhai P, Zhang Y, et al. The *Aspergillus fumigatus* damage resistance protein family coordinately regulates ergosterol biosynthesis and azole susceptibility[J]. *mBio*, 2016, 7(1):e01919-15.
- [55] Liu FF, Pu L, Zheng QQ, et al. Calcium signaling mediates antifungal activity of triazole drugs in the *Aspergilli*[J]. *Fungal Genet Biol*, 2015(81):182-190.
- [56] Gonzalez-Jimenez I, Lucio J, Amich J, et al. A cyp51B mutation contributes to azole resistance in *Aspergillus fumigatus*[J]. *J Fungi (Basel, Switzerland)*, 2020, 6(4):315.
- [57] Gonzalez-Jimenez I, Lucio J, Roldan A, et al. Are point mutations in HMG-CoA reductases (Hmg1 and Hmg2) a step towards azole resistance in *Aspergillus fumigatus*[J]. *Molecules*, 2021, 26(19):5975.
- [58] Ramage G, Rajendran R, Sherry L, et al. Fungal biofilm resistance[J]. *Int J Microbiol*, 2012(2012):528521.
- [59] Chen M, Zhong G, Wang S, et al. Deletion of cox7c results in Pan-Azole resistance in *Aspergillus fumigatus*[J]. *Antimicrob Agents Chemother*, 2022, 66(6):e0015122.
- [60] Kamiński DM. Recent progress in the study of the interactions of amphotericin B with cholesterol and ergosterol in lipid environments[J]. *Eur Biophys J*, 2014, 43(10-11):453-467.
- [61] 王爱平, 李若瑜. 系统抗真菌药物概述[J]. 中国药物评价, 2012, 29(1):10-13.
- [62] Shekhova E, Kniemeyer O, Brakhage AA. Induction of mitochondrial reactive oxygen species production by itraconazole, terbinafine, and amphotericin B as a mode of action against *Asper-*

- gillus fumigatus*[J]. Antimicrob Agents Chemother, 2017, 61(11):e00978-17.
- [63] Posch W, Blatzer M, Wilflingseder D, et al. *Aspergillus terreus*: Novel lessons learned on amphotericin B resistance[J]. Med Mycol, 2018, 56(suppl_1):73-82.
- [64] Reichert-Lima F, Lyra L, Pontes L, et al. Surveillance for azoles resistance in *Aspergillus* spp. highlights a high number of amphotericin B-resistant isolates[J]. Mycoses, 2018, 61(6):360-365.
- [65] Ashu EE, Korfanty GA, Samarasinghe H, et al. Widespread amphotericin B-resistant strains of *Aspergillus fumigatus* in Hamilton, Canada[J]. Infect Drug Resist, 2018(11):1549-1555.
- [66] Fan Y, Korfanty GA, Xu J. Genetic Analyses of amphotericin B susceptibility in *Aspergillus fumigatus*[J]. J Fungi (Basel), 2021, 7(10):860.
- [67] Gow N, Latge JP, Munro CA. The Fungal cell wall: structure, biosynthesis, and function[J]. Microbiol Spectr, 2017, 5(3):0035.
- [68] Garcia-Rubio R, de Oliveira HC, Rivera J, et al. The fungal cell wall: candida, cryptococcus, and aspergillus species[J]. Front Microbiol, 2019, 10:2993.
- [69] Liu W, Yuan L, Wang S. Recent progress in the discovery of antifungal agents targeting the cell Wall[J]. J Med Chem, 2020, 63(21):12429-12459.
- [70] Aguilar-Zapata D, Petraitene R, Petraitis V. Echinocandins: the expanding antifungal armamentarium[J]. Clin Infect Dis, 2015, 61(Suppl 6):S604-611.
- [71] Sharma C, Chowdhary A. Molecular bases of antifungal resistance in filamentous fungi[J]. Int J Antimicrob Agents, 2017, 50(5):607-616.
- [72] Gardiner RE, Souteropoulos P, Park S, et al. Characterization of *Aspergillus fumigatus* mutants with reduced susceptibility to caspofungin[J]. Med Mycol, 2005, 43 Suppl 1:S299-305.
- [73] Rocha EM, Garcia-Effron G, Park S, et al. A Ser678Pro substitution in Fks1p confers resistance to echinocandin drugs in *Aspergillus fumigatus*[J]. Antimicrob Agents Chemother, 2007, 51(11):4174-4176.
- [74] Steinbach WJ, Lamothe F, Juvvadi PR. Potential microbiological effects of higher dosing of echinocandins[J]. Clin Infect Dis, 2015, 61(Suppl 6):S669-677.
- [75] Dichtl K, Ebel F, Dirr F, et al. Farnesol misplaces tip-localized Rho proteins and inhibits cell wall integrity signalling in *Aspergillus fumigatus*[J]. Mol Microbiol, 2010, 76(5):1191-1204.
- [76] Moreno-Vel squez SD, Seidel C, Juvvadi PR, et al. Caspofungin-mediated growth inhibition and paradoxical growth in *aspergillus fumigatus* involve fungicidal hyphal tip lysis coupled with regenerative intrahyphal growth and dynamic changes in β -1,3-glucan synthase localization[J]. Antimicrob Agents Chemother, 2017, 61(10):e00710-17.
- [77] Kerkaert JD, Le Mauff F, Wucher BR, et al. An Alanine aminotransferase is required for biofilm-specific resistance of *Aspergillus fumigatus* to echinocandin treatment[J]. mBio, 2022, 13(2):e0293321.
- [78] Chen Y, Le Mauff F, Wang Y, et al. The transcription factor so-ma synchronously regulates biofilm formation and cell wall homeostasis in *Aspergillus fumigatus*[J]. mBio, 2020, 11(6):e02329-20.
- [79] Zhang H, Zhou W, Zhang W, et al. inhibitory effects of citral, cinnamaldehyde, and tea polyphenols on mixed biofilm formation by foodborne *Staphylococcus aureus* and *Salmonella enteritidis*[J]. J Food Prot, 2014, 77(6):927-933.
- [80] Khan MS, Ahmad I. Antibiofilm activity of certain phytocompounds and their synergy with fluconazole against *Candida albicans* biofilms[J]. J Antimicrob Chemother, 2012, 67(3):618-21.
- [81] 黄宏,陈一强,孔晋亮,等.不同中药单体成分对两性霉素B耐药的烟曲霉抑菌活性的体外研究[J].中国现代医药杂志,2014,16(2):1-4.
- [82] 邬丽红,陈一强,孔晋亮,等.肉桂醛对烟曲霉菌体外生物膜的影响[J].重庆医学,2016,45(3):326-328.
- [83] Didehdar M, Chegini Z, Tabaeian SP, et al. Cinnamomum: The new therapeutic agents for inhibition of bacterial and fungal biofilm-associated infection[J]. Front Cell Infect Microbiol, 2022, 12:930624.
- [84] 龙凯,谢小梅,方建如,等.肉桂醛、柠檬醛对烟曲霉色素及关键基因 alb1 mRNA 表达的影响[J].微生物学通报,2007,7(3):541-544.
- [85] 罗闵丹,杨芬,施旻,等.柠檬醛对小鼠侵袭性肺曲霉病的作用[J].中草药,2009,40(3):428-430.
- [86] Davis LE, Shen JK, Cai Y. Antifungal activity in human cerebrospinal fluid and plasma after intravenous administration of Allium sativum[J]. Antimicrob Agents Chemother, 1990, 34(4):651-653.
- [87] Yamada Y, Azuma K. Evaluation of the *in vitro* antifungal activity of allicin[J]. Antimicrob Agents Chemother, 1977, 11(4):743-749.
- [88] Shadkhan Y, Shemesh E, Mirelman D, et al. Efficacy of allicin, the reactive molecule of garlic, in inhibiting *Aspergillus* spp. in vitro, and in a murine model of disseminated aspergillosis[J]. J Antimicrob Chemother, 2004, 53(5):832-836.
- [89] Schier C, Foerster N e Reiter J, Heupel M, et al. Allicin as a volatile or nebulisable antimycotic for the treatment of pulmonary mycoses: *in vitro* studies using a lung flow test Rig[J]. Int J Mol Sci, 2022, 23(12):6607.
- [90] Freeman F, Kodera, Y. Garlic chemistry; stability of S-(2-propenyl)-2-propene-1-sulfinothioate (allicin) in blood, solvents, and simulated physiological fluids[J]. J Agricul Food Chem, 1995, 43 (9):2332-2338.
- [91] 高磊.天然药物小檗碱抑制烟曲霉作用机制的研究[D].长春:吉林大学,2012.

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